

Use of CSAIDs in Rhinovirus infection

5

Field of Invention

The present invention relates to the use of a CSBP/p38 inhibitor in the treatment of a CSBP/p38 mediated disease.

10 Background of the Invention

Human rhinovirus (HRV), the most frequent cause of the common cold, is increasingly associated with more serious sequelae including exacerbation's of asthma, chronic bronchitis, COPD, otitis media, and sinusitis (*Gern et al.*, Clin Micro Reviews 12(1): 9-18 (1999); *Pitkaranta, and Hayden*, Annals of Medicine 30 (6): 529-537 (1998); *Seemungal et al*, ATS abstract "Rhinoviruses are associated with exacerbation's of COPD" (1998)). Recent published studies in adults and adolescents, using PCR to assist in viral detection, have shown that up to 50 to 80% of asthma exacerbation's are associated with upper respiratory tract virus infection, and that rhinovirus is the most common virus (*Atmar et al*, Archives of Internal Medicine. 158 (22): 2453-9 (1998); *Johnston, SL.*, British Medical Journal 310: 1225-9 (1995)). HRV infects nasal epithelial cells; recent evidence suggests the virus may also infect bronchial epithelium. Prodromal cold symptoms are apparent within 24 hours post-infection, peak on days 2 through 5, and resolve within seven to fourteen days; but can be more protracted in some individuals. Symptoms are believed to arise more from the host's response to infection, than an acute cytotoxic effect, since only a small fraction of upper respiratory epithelial cells are demonstrably infected, and there is minimal epithelial cell damage (*Winther et al*, JAMA 256: 1763-1767 (1986). Increased intranasal levels of kinins, IL-1, IL-8, IL-6, IL-11, and neutrophils are found in normal individuals infected with rhinoviruses. A correlation between IL-8 concentration in nasal secretions with local myeloperoxidase levels and with symptom severity has been demonstrated in several recent studies (*Grieff, et al*. Eur Respir J 13: 41-47 (1999); *Teren, et al*. Am J Respir Crit Care Med 155: 1362-1366 (1997), *Turner, et al*. Clin Infect Dis 26: 840-846 (1998). Intranasal concentrations of IL-1 and IL-6 have been correlated with symptom severity as well (*Proud et al*, J. Infect. Dis. 169:1007-1013 (1994); *Zhu et al*, J. Clin. Invest. 97:421-430 (1996)). Experimental rhinovirus infection also results in enhanced immediate and late phase allergic reactions, and in increased infiltration of T lymphocytes and eosinophils into the lower airways. In atopics and

WO 01/19322

PCT/US00/25386

asthmatics, these effects persist for up to 2 months post - infection (*Gern and Busse*, Am J Respir Crit Care Med, 152: S40-S45 (1995). Human bronchial epithelial cell lines have been shown to produce IL-1, IL-6, IL-8, IL-11 and GM-CSF in response to rhinovirus infection (*Subauste et al*, J Clin Invest, 96: 549-557 (1995); *Gern et al.*, *supra*, 1999). Early production of cytokines by rhinovirus - infected epithelial cells may therefore be responsible for triggering recruitment of neutrophils, T cells and activated eosinophils into the upper and lower airways.

In addition, IL-1, IL-6, and IL-8 are also produced in response to infection with other respiratory viruses (influenza, respiratory syncytial virus) which can cause the common cold and associated sequelae.

By interfering with the biochemical processes of epithelial cells resulting from virus infection there represents a viable new therapeutic target by an inhibitor of CSBP/p38. This invention is directed to the novel discovery of treatment of this therapeutic target.

15

Summary of the Invention

The present invention relates to the use of a CSBP/p38 kinase inhibitor for the treatment, including prophylaxis, of the common cold, or respiratory viral infection caused by human rhinovirus infection (HRV), other enteroviruses, coronavirus, influenza virus, parainfluenza virus, respiratory syncytial virus, or adenovirus infection in a human in need thereof which method comprises administering to said human an effective amount of a CBSP/p38 inhibitor.

Another aspect of the present invention is a method of treating, including prophylaxis of influenza induced pneumonia in a human in need thereof which method comprises administering to said human an effective amount of a CBSP/p38 inhibitor

The present invention also relates to the use of the CSBP/p38 kinase inhibitor for the treatment, including prophylaxis, of inflammation associated with a viral infection of a human rhinovirus (HRV), other enteroviruses, coronavirus, influenza virus, parainfluenza virus, respiratory syncytial virus, or adenovirus.

Brief Description of the Drawings

Figure 1 demonstrates Cytokine Production by Rhinovirus infected BEAS-2B cells. Culture supernatants were collected 72 hours post-infection of BEAS-2B cells with rhinovirus-39 (MOI 1). Uninfected cells served as controls. Protein concentrations in supernatants were determined by ELISA (R&D Systems). Results represent the mean concentration values obtained from 6 experiments.

Figure 2 demonstrates Inhibition of cytokines by CSAIDs: BEAS-2B cultures infected with rhinovirus-39 were cultured in the presence of various concentrations of drug. Cytokine levels in supernatant were determined 72 hours post-infection using commercially available ELISA kits. Results are expressed as % inhibition from infected untreated cultures. Cytokine concentrations in infected control cultures were 4902 pg/ml IL-6, 4520 pg/ml IL-8, and 28 pg/ml GM-CSF.

Figure 3 demonstrates Tyrosine phosphorylation of p38 kinase by rhinovirus infection. BEAS-2B cells were incubated with rhinovirus-39 for various times as indicated. Cell lysates were separated by 10% SDS-polyacrylamide gel, transferred to nitrocellulose membrane and probed with specific antibody to phosphorylated p38 kinase (A) or total p38 kinase (B). Amounts of p38 kinase were quantitated by image analyzer and are presented as volumes based on densitometer scans.

Figure 4 demonstrates Tyrosine phosphorylation of p38 kinase by rhinovirus infection. BEAS-2B cells were incubated with various doses (MOI) of rhinovirus-39 for 30 minutes. Cell lysates were separated by 10% SDS-polyacrylamide gel, transferred to nitrocellulose membrane and probed with specific antibody to phosphorylated p38 kinase or total p38 kinase. Amounts of p38 kinase were quantitated by image analyzer and are presented as relative amounts of total or phosphorylated p38 kinase compared to control cells incubated with media alone (fold increase).

Figure 5 demonstrates the effect of Compound VI, 1-(1,3-Dihydroxyprop-2-yl)-4-(4-fluorophenyl)-5-[2-phenoxy-pyrimidin-4-yl]imidazole on improvement of pulmonary function with increasing doses. BALB/c mice were treated from days 3-8 post-infection with a sub-lethal dose of Influenza A. Pulmonary resistance was determined using whole body plethysmography.

Figure 6 demonstrates the effect of Compounds V, 1-(4-Piperidinyl)-4-(4-fluorophenyl)-5-(2-methoxy-4-pyrimidinyl)imidazole and Compound VI, on prevention of weight loss in animals in an in vivo influenza model.

Figure 7 demonstrates the efficacy of of Compounds V and VI at improving arterial blood oxygen levels (%SpO₂) upon treatment. SpO₂ was determined using daily pulse oximetry.

Detailed Description of the Invention

IL-1, TNF, and other cytokines affect a wide variety of cells and tissues and these cytokines as well as other leukocyte derived cytokines are important and critical inflammatory mediators of a wide variety of disease states and conditions.

WO 01/19322

PCT/US00/25386

The inhibition of these cytokines is of benefit in controlling, reducing and alleviating many of these disease states.

In particular, the present invention is directed to the treatment of a viral infection in a human, which is caused by the human rhinovirus (HRV), other
5 enterovirus, coronavirus, influenza virus, parainfluenza virus, respiratory syncytial virus, or an adenovirus. In particular the invention is directed to respiratory viral infections that exacerbate asthma (induced by such infections), chronic bronchitis, chronic obstructive pulmonary disease, otitis media, and sinusitis. While inhibiting IL-8 or other cytokines may be beneficial in treating a rhinovirus may be known,
10 the use of an inhibitor of the p38 kinase for treating HRV or other respiratory viral infections causing the common cold is believed novel.

It should be noted that the respiratory viral infection treated herein may also be associated with a secondary bacterial infection, such as otitis media, sinusitis, or pneumonia.

15 For use herein treatment may include prophylaxis for use in a treatment group susceptible to such infections. It may also include reducing the symptoms of, ameliorating the symptoms of, reducing the severity of, reducing the incidence of, or any other change in the condition of the patient, which improves the therapeutic outcome.

20 The mechanism of action for inhibition of a cytokine by a cytokine suppressive anti-inflammatory drug (CSAID) versus inhibition of virus - induced IL-8 production in airway epithelial cells is believed to be different. In the rhinovirus system, IL-8 production, and CSAID inhibition of IL-8 synthesis is independent of IL-1 and TNF production, whereas the published studies have
25 focused on IL-1 and TNF - induced IL-8 production.

It should be noted that the treatment herein is not directed to the elimination or treatment of the viral organism itself but is directed to treatment of the respiratory viral infection that exacerbates other diseases or symptoms of disease, such as asthma (induced by such infections), chronic bronchitis, chronic obstructive pulmonary disease, otitis
30 media, and sinusitis.

The present invention will demonstrate that CSAID inhibitors are useful in the treatment of symptoms associated with HRV, including exacerbations of underlying conditions such as asthma, COPD, sinusitis and otitis media amongst others.

35 A preferred virus for treatment herein is the human rhinovirus infection (HRV) or respiratory syncytial virus (RSV).

Another aspect of the present invention is a method of treating, including prophylaxis of influenza induced pneumonia in a human in need thereof which method comprises administering to said human an effective amount of a CBSP/p38 inhibitor. Therefore, for this usage, a preferred virus for treatment is the influenza virus.

Lastly, another aspect of the present invention relates to the use of a CSBP/p38 kinase inhibitor for the treatment, including prophylaxis, of inflammation associated with a viral infection of a human rhinovirus (HRV), other enteroviruses, coronavirus, influenza virus, parainfluenza virus, respiratory syncytial virus, or adenovirus. Preferably the viral infection is HRV or RSV, or the influenza or parainfluenza virus.

Suitable CSAID compounds are well known in the art, and an assay for determining CBSP/p38 inhibition is also readily available using assays disclosed in the below noted patents or applications. For instance, see US Patents 5,716,972, US 5,686,455, US 5,656,644, US 5,593,992, US 5,593,991, US 5,663,334, US 5,670,527, US 5,559,137, 5,658,903, US 5,739,143, US 5,756,499, and US 5,716,955; WIPO publications WO 98/25619, WO 97/25048, WO 99/01452, WO 97/25047, WO 99/01131, WO 99/01130, WO 97/33883, WO 97/35856, WO 97/35855, WO 98/06715, WO 98/07425, WO 98/28292, WO 98/56377, WO 98/07966, WO 99/01136, WO 99/17776, WO 99/01131, WO 99/01130, WO 99/32121, WO 00/26209, WO 99/58502, WO 99/58523, WO 99/57101, WO 99/61426, WO 99/59960, WO 99/59959, WO 00/18738, WO 00/17175, WO 99/17204, WO 00/20402, WO 99/64400, WO 00/01688, WO 00/07980, WO 00/07991, WO 00/06563, WO 00/12074, WO 00/12497, WO 00/31072, WO 00/31063, WO 00/23072, WO 00/31065, WO 00/35911, WO 00/39116, WO 00/43384, WO 00/41698, WO 97/36587, WO 97/47618, WO 97/16442, WO 97/16441, WO 97/12876, WO 98/7966, WO 98/56377, WO 98/22109, WO 98/24782, WO 98/24780, WO 98/22457, WO 98/52558, WO 98/52941, WO 98/52937, WO 98/52940, WO 98/56788, WO 98/27098, WO 99/00357, WO 98/47892, WO 98/47899, WO 99/03837, WO 99/01441, WO 99/01449, WO 99/03484, WO 95/09853, WO 99/15164, WO 98/50356, WO 95/09851, WO 95/09847, WO 95/09852, WO 92/12154, WO 94/19350, DE 19842833, JP 2000 86657 and De Laszlo et al., Bioorg. Med. Chem. Lett 8 (1998) 2689-2694 whose disclosures are all incorporated herein by reference in their entirety.

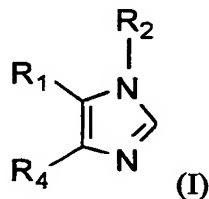
Preferred compounds of this invention include those contained in WO 99/01131, and a representative genus is described below. Also preferred for use herein are the compounds disclosed in WO 99/61426 Scios, Inc.; and those compounds disclosed in WO 98/27098 containing the compound known as VX-745; (also known as 5-(2,6-Dichloro-phenyl)-2-(2,4-difluoro-phenylsulfanyl)-1,7,8a-

WO 01/19322

PCT/US00/25386

triazza-naphthalen-6-one), the Johnson & Johnson compound RWJ-68354 disclosed in WO 98/47899, RPR compound RPR-200765A, the Zeneca compound ZM 336372 disclosed in WO 99/15164; the Sugen compound SU 4984 disclosed in WO 98/50356. A review of various inhibitors of p38 kinase is taught in Boehm et al.,
 5 Exp. Opin. Ther. Patents 10(1):25-37 (2000).

Compounds of Formula (I) are represented by the formula:



10 wherein

R₁ is 4-pyridyl, pyrimidinyl, 4-pyridazinyl, 1,2,4-triazin-5-yl, quinolyl, isoquinolinyl, or quinazolin-4-yl ring, which ring is substituted with Y-R_a and optionally with an additional independent substituent selected from C₁₋₄ alkyl, halogen, hydroxyl, C₁₋₄ alkoxy, C₁₋₄ alkylthio, C₁₋₄ alkylsulfinyl, CH₂OR₁₂,
 15 amino, mono and di- C₁₋₆ alkyl substituted amino, an N-heterocyclyl ring which ring has from 5 to 7 members and optionally contains an additional heteroatom selected from oxygen, sulfur or NR₁₅, N(R₁₀)C(O)R_b or NHR_a;

Y is oxygen or sulfur;

R₄ is phenyl, naphth-1-yl or naphth-2-yl, or a heteroaryl, which is optionally
 20 substituted by one or two substituents, each of which is independently selected, and which, for a 4-phenyl, 4-naphth-1-yl, 5-naphth-2-yl or 6-naphth-2-yl substituent, is halogen, cyano, nitro, C(Z)NR₇R₁₇, C(Z)OR₁₆, (CR₁₀R₂₀)_vCOR₁₂, SR₅, SOR₅, OR₁₂, halo-substituted-C₁₋₄ alkyl, C₁₋₄ alkyl, ZC(Z)R₁₂, NR₁₀C(Z)R₁₆, or (CR₁₀R₂₀)_vNR₁₀R₂₀ and which, for other
 25 positions of substitution, is halogen, cyano, C(Z)NR₁₃R₁₄, C(Z)OR₃, (CR₁₀R₂₀)_m"COR₃, S(O)_mR₃, OR₃, halo-substituted-C₁₋₄ alkyl, C₁₋₄ alkyl, (CR₁₀R₂₀)_m"NR₁₀C(Z)R₃, NR₁₀S(O)_mR₈, NR₁₀S(O)_m"NR₇R₁₇, ZC(Z)R₃ or (CR₁₀R₂₀)_m"NR₁₃R₁₄;

Z is oxygen or sulfur;

30 n is an integer having a value of 1 to 10;

m is 0, or the integer 1 or 2;

m' is an integer having a value of 1 or 2,

m" is 0, or an integer having a value of 1 to 5;

WO 01/19322

PCT/US00/25386

v is 0, or an integer having a value of 1 or 2;

R₂ is -C(H) (A) (R₂₂);

A is an optionally substituted aryl, heterocyclyl, or heteroaryl ring, or A is a substituted C₁₋₁₀ alkyl;

5 R₂₂ is an optionally substituted C₁₋₁₀ alkyl;

R_a is aryl, arylC₁₋₆alkyl, heterocyclic, heterocyclylC₁₋₆ alkyl, heteroaryl, heteroarylC₁₋₆alkyl, wherein each of these moieties may be optionally substituted;

10 R_b is hydrogen, C₁₋₆ alkyl, C₃₋₇ cycloalkyl, aryl, arylC₁₋₄ alkyl, heteroaryl, heteroarylC₁₋₄alkyl, heterocyclyl, or heterocyclylC₁₋₄ alkyl, wherein each of these moieties may be optionally substituted;

R₃ is heterocyclyl, heterocyclylC₁₋₁₀ alkyl or R₈;

R₅ is hydrogen, C₁₋₄ alkyl, C₂₋₄ alkenyl, C₂₋₄ alkynyl or NR₇R₁₇, excluding the moieties SR₅ being SNR₇R₁₇ and SOR₅ being SOH;

15 R₆ is hydrogen, a pharmaceutically acceptable cation, C₁₋₁₀ alkyl, C₃₋₇ cycloalkyl, aryl, arylC₁₋₄ alkyl, heteroaryl, heteroarylC₁₋₄alkyl, heterocyclyl, aroyl, or C₁₋₁₀ alkanoyl;

20 R₇ and R₁₇ is each independently selected from hydrogen or C₁₋₄ alkyl or R₇ and R₁₇ together with the nitrogen to which they are attached form a heterocyclic ring of 5 to 7 members which ring optionally contains an additional heteroatom selected from oxygen, sulfur or NR₁₅;

25 R₈ is C₁₋₁₀ alkyl, halo-substituted C₁₋₁₀ alkyl, C₂₋₁₀ alkenyl, C₂₋₁₀ alkynyl, C₃₋₇ cycloalkyl, C₅₋₇ cycloalkenyl, aryl, arylC₁₋₁₀ alkyl, heteroaryl, heteroarylC₁₋₁₀ alkyl, (CR₁₀R₂₀)_nOR₁₁, (CR₁₀R₂₀)_nS(O)_mR₁₈, (CR₁₀R₂₀)_nNHS(O)₂R₁₈, (CR₁₀R₂₀)_nNR₁₃R₁₄; wherein the aryl, arylalkyl, heteroaryl, heteroaryl alkyl may be optionally substituted;

R₉ is hydrogen, C(Z)R₁₁ or optionally substituted C₁₋₁₀ alkyl, S(O)₂R₁₈, optionally substituted aryl or optionally substituted aryl-C₁₋₄ alkyl;

R₁₀ and R₂₀ is each independently selected from hydrogen or C₁₋₄ alkyl;

30 R₁₁ is hydrogen, C₁₋₁₀ alkyl, C₃₋₇ cycloalkyl, heterocyclyl, heterocyclyl C₁₋₁₀alkyl, aryl, arylC₁₋₁₀ alkyl, heteroaryl or heteroarylC₁₋₁₀ alkyl, wherein these moieties may be optionally substituted;

R₁₂ is hydrogen or R₁₆;

35 R₁₃ and R₁₄ is each independently selected from hydrogen or optionally substituted C₁₋₄ alkyl, optionally substituted aryl or optionally substituted aryl-C₁₋₄ alkyl, or together with the nitrogen which they are attached form a heterocyclic ring of

5 to 7 members which ring optionally contains an additional heteroatom selected from oxygen, sulfur or NR₉ ;

R₁₅ is R₁₀ or C(Z)-C₁₋₄ alkyl;

R₁₆ is C₁₋₄ alkyl, halo-substituted-C₁₋₄ alkyl, or C₃₋₇ cycloalkyl;

- 5 R₁₈ is C₁₋₁₀ alkyl, C₃₋₇ cycloalkyl, heterocyclyl, aryl, aryl₁₋₁₀alkyl, heterocyclyl, heterocyclyl-C₁₋₁₀alkyl, heteroaryl or heteroaryl₁₋₁₀alkyl; or a pharmaceutically acceptable salt thereof.

R₂ is a substituted alkyl derivative. It is recognized that the first methylene carbon in this chain is a tertiary carbon, and it will contain one hydrogen moiety. This methylene group will have has two additional substituents, an R₂₂ moiety and an A moiety, -C(H)(A)(R₂₂). Both A and R₂₂ may not be unsubstituted C₁₋₁₀ alkyl moieties.

In a preferred embodiment, R₂ is a -C(AA₁)(A) moiety, wherein AA₁ is the R₂₂ moiety, but is specifically the side chain residue (R) of an amino acid, as is further described herein.

Suitably, A is an optionally substituted C₃₋₇cycloalkyl, aryl, heteroaryl, or heterocyclic ring, or A is a substituted C₁₋₁₀ alkyl moiety.

When A is an aryl, heteroaryl and heterocyclic ring, the ring may be substituted independently one or more times, preferably, 1 to 3 times by C₁₋₁₀ alkyl; halogen; halo substituted C₁₋₁₀ alkyl, such as CF₃; (CR₁₀R₂₀)_tOR₁₁; (CR₁₀R₂₀)_tNR₁₃R₁₄, especially amino or mono- or di-C₁₋₄ alkylamino; (CR₁₀R₂₀)_tS(O)_mR₁₈, wherein m is 0, 1 or 2; SH; NR₁₀C(Z)R₃ (such NHCO(C₁₋₁₀ alkyl)); or NR₁₀S(O)_mR₈ (such as NHSO₂(C₁₋₁₀ alkyl)).

Suitably, t is 0, or an integer of 1 to 4.

When A is an optionally substituted cycloalkyl it is as defined below with the R₂₂ substitution.

When A is an optionally substituted heterocyclyl ring, the ring is preferably a morpholino, pyrrolidinyl, piperazinyl or a piperidinyl ring.

When A is an optionally substituted aryl moiety, it is preferably a phenyl ring.

When A is an optionally substituted heteroaryl ring, it is as defined below in the definition section.

When A is a substituted C₁₋₁₀ alkyl moiety, the alkyl chain may be straight or branched. The chain is substituted independently 1 or more times, preferably 1 to 3 times by halogen, such as fluorine, chlorine, bromine or iodine; halosubstituted C₁₋₁₀ alkyl, such as CF₃; C₃₋₇cycloalkyl, C₁₋₁₀ alkoxy, such as methoxy or ethoxy; hydroxy substituted C₁₋₁₀ alkoxy; halosubstituted C₁₋₁₀ alkoxy, such as OCF₂CF₂H;

WO 01/19322

PCT/US00/25386

OR₁₁; S(O)_mR₁₈ (wherein m is 0, 1 or 2); NR₁₃R₁₄; C(Z)NR₁₃R₁₄;
 S(O)_mNR₁₃R₁₄; NR₂₃C(Z)R₁₁; NHS(O)₂R₁₈; C(Z)R₁₁; OC(Z)R₁₁; C(Z)OR₁₁;
 C(Z)NR₁₁OR₉; N(OR₆)C(Z)NR₁₃R₁₄; N(OR₆)C(Z)R₁₁; C(=NOR₆)R₁₁;
 NR₂₃C(=NR₁₉)NR₁₃R₁₄; OC(Z)NR₁₃R₁₄; NR₂₃C(Z)NR₁₃R₁₄; or

5 NR₂₃C(Z)OR₁₀.

Preferably A is a C₃₋₇ cycloalkyl, or a C₁₋₆ alkyl, more preferably a C₁₋₂ alkyl, i.e. a methylene or ethylene moiety, more preferably a methylene moiety which is substituted by one of the above noted groups.

Preferably, when A is a C₁₋₁₀ alkyl, it is substituted by OR₁₁ where R₁₁ is
 10 preferably hydrogen, aryl or arylalkyl; NR₁₃R₁₄; OC(Z)R₁₁; or C(Z)OR₁₁.

More preferably, A is substituted by OR₁₁ where R₁₁ is hydrogen.

Suitably, R₂₂ is a C₁₋₁₀ alkyl chain, which chain may be straight or branched and which may be optionally substituted independently, one or more times, preferably 1 to 3 times, by halogen, such as fluorine, chlorine, bromine or iodine; halo substituted
 15 C₁₋₁₀ alkyl; C₁₋₁₀ alkoxy, such as methoxy or ethoxy; hydroxy substituted C₁₋₁₀ alkoxy; halosubstituted C₁₋₁₀ alkoxy, such as OCF₂CF₂H; OR₁₁; S(O)_mR₁₈;
 NR₁₃R₁₄; C(Z)NR₁₃R₁₄; S(O)_mNR₁₃R₁₄; NR₂₃C(Z)R₁₁; NHS(O)₂R₁₈;
 C(Z)R₁₁; OC(Z)R₁₁; C(Z)OR₁₁; C(Z)NR₁₁OR₉; N(OR₆)C(Z)NR₁₃R₁₄;
 N(OR₆)C(Z)R₁₁; C(=NOR₆)R₁₁; NR₂₃C(=NR₁₉)NR₁₃R₁₄; OC(Z)NR₁₃R₁₄;
 20 NR₂₃C(Z)NR₁₃R₁₄; NR₂₃C(Z)OR₁₀; optionally substituted C₃₋₇ cycloalkyl;
 optionally substituted aryl, such as phenyl; optionally substituted heteroaryl; or an optionally substituted heterocyclic. The optional substituents on these cycloalkyl, aryl, heteroaryl, and heterocyclic moieties are as defined herein below.

It is noted that those R₂₂ substituent groups which contain carbon as the first
 25 connecting group, i.e. C(Z)OR₁₁; C(Z)NR₁₁OR₉; C(Z)R₁₁; C(Z)NR₁₃R₁₄, and C(=NOR₆)R₁₁, may be the sole carbon in alkyl chain. Therefore, the R₂₂ group may, for instance, be a carboxy, an aldehyde, or an amide, as well as being a substituent off a methylene unit, such as carbamoylmethyl, or acetamidomethyl.
 Preferably R₂₂ is a C₁₋₆ unsubstituted or substituted alkyl group, such as a C₁₋₃
 30 alkylene, such as methyl, ethyl or isopropyl, or a methylene or ethylene moiety substituted by one of the above noted moieties, or as noted above those substituent groups which contain a carbon may substituent for the first methylene unit of the alkyl chain, such as carboxy, C(O)OR₁₁, C(O)NR₁₃R₁₄, or R₂₂ is an optionally substituted aryl group, such as a benzyl or phenethyl. In other words, R₂₂ can be an optionally
 35 substituted alkyl group, or R₂₂ can be C(Z)OR₁₁, C(Z)NR₁₁OR₉, C(Z)R₁₁, C(Z)NR₁₃R₁₄, or C(=NOR₆)R₁₁.

WO 01/19322

PCT/US00/25386

Preferably R₂₂ is a C₁₋₆ unsubstituted or substituted alkyl group, more preferably a C₁₋₂ alkylene chain, such as a methylene or ethylene moiety, more preferably methylene.

Preferably the alkyl chain is substituted by OR₁₁, where R₁₁ is preferably hydrogen, aryl or arylalkyl; S(O)mR₁₈, where m is 0 and R₁₈ is a C₁₋₆ alkyl; or an optionally substituted aryl, i.e. a benzyl or phenethyl moiety.

More preferably, R₂₂ is phenyl, benzyl, CH₂OH, or CH₂-O-aryl.

Preferably, one or both of A and R₂₂ contain hydroxy moieties, such as in C₁₋₆ alkyl OR₁₁, wherein R₁₁ is hydrogen, i.e. CH₂CH₂OH.

Suitably, when AA₁ is the (R) side chain residue of an amino acid, it is a C₁₋₆ alkyl group, which may be straight or branched. This means the R group off the core amino acid of the structure R-C(H)(COOH)(NH₂). The R residue term is for example, CH₃ for alanine, (CH₃)₂CH- for valine, (CH₃)₂CH-CH₂- for leucine, phenyl-CH₂- for phenylalanine, CH₃-S-CH₂-CH₂- for methionine, etc. All generally recognized primary amino acids are included in this groups, such as but not limited to, alanine, arginine, asparagine, aspartic acid, cysteine, glutamine, glutamic acid, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, serine, threonine, tryptophan, tyrosine, valine, hydroxylysine, methylhistidine, and other naturally occurring amino acids not found in proteins, such as β-alanine, γ-aminobutyric acid, homocysteine, homoserine, citrulline, ornithine, canavanine, djenkolic acid, and β-cyanoalanine, or other naturally occurring non-mammalian amino acids.

Preferably AA₁ is the residue of phenylalanine, or alanine.

Preferably, A is a hydroxy substituted C₁₋₁₀ alkyl, and R₂₂ is a C₁₋₁₀ alkyl or a hydroxy substituted C₁₋₁₀ alkyl.

For further definitions please refer to the descriptions in WO 99/01131, or in WO 99/01136, *supra*.

A preferred compound for use of 1-(1,3-Dihydroxyprop-2-yl)-4-(4-fluorophenyl)-5-(2-phenoxy pyrimidin-4-yl)imidazole, or a pharmaceutically acceptable salt thereof.

Other suitable compounds for use herein include but are not limited to, *trans*-1-(4-Hydroxycyclohexyl)-4-(4-fluorophenyl)-5-[(2-methoxy)pyrimidin-4-yl]imidazole; 1-(4-Piperidiny)-4-(4-fluorophenyl)-5-(2-methoxy-4-pyrimidinyl)imidazole; or (4-Fluorophenyl)-2-(4-methylsulfinylphenyl)-5-(4-pyridyl)-imidazole.

Methods of using and dosage amounts are the same as those disclosed in the references cited above. See for instance, Adams et al., US patent 5,756,499, issued 26 May 1998. In order to use a compound of formula (I) or a pharmaceutically

acceptable salt thereof in therapy, it will normally be formulated into a pharmaceutical composition in accordance with standard pharmaceutical practice.

For all methods of use disclosed herein (or the compounds of Formula (I) and other CSAID compounds), suitably, the daily oral dosage regimen will be from
5 about 0.1 to about 80 mg/kg of total body weight, preferably from about 0.2 to 30 mg/kg, more preferably from about 0.5 mg to 15mg. The daily parenteral dosage regimen about 0.1 to about 80 mg/kg of total body weight, preferably from about 0.2 to about 30 mg/kg, and more preferably from about 0.5 mg to 15mg/kg. The daily
10 topical dosage regimen will preferably be from 0.1 mg to 150 mg, administered one to four, preferably two or three times daily. The daily inhalation dosage regimen will preferably be from about 0.01 mg/kg to about 1 mg/kg per day.

The novel use of CSAID compounds herein may also be used in association with the veterinary treatment of mammals, other than humans, in need of inhibition of CSBP/p38 or cytokine inhibition or production for treatment of influenza
15 pneumonia, and other sequelae associated with viral infection.

The CSBP/p38 inhibitor may also be administered with a second therapeutic agent. The second therapeutic agent may be an antiviral agent such as ribavirin, amantidine, rimantidine, Pleconaril, AG 7088 or BTA-188; it may also be an
20 antiviral agent such as an influenza neuraminidase inhibitor, such as zanamivir (Relenza), oseltamivir (Tamiflu) or RWJ-270201; it may be an antihistamine, such as Benadryl®, chlorpheniramine and salts thereof, brompheniramine or salts thereof, and the generally accepted non-sedating antihistamines, such as loratadine (Claritin®), descarboethoxyloratadine (DCL), fexofenadine (Allegra®), and
25 cetirizine hydrochloride (Zyrtec®) etc., a decongestant, such as phenylpropanolamine and salts thereof, pseudoephedrine or salts thereof; steroids, such as dexamethasone, prednisone, or prednisolone, etc.; various antibiotics, such as the quinolones, cephalosporins, β -lactamase inhibitors, etc.; anti-inflammatory agents, such as an NSAID, a COX-1 or COX-2 inhibitor, ASA, or indomethacin, etc. It is recognized that the above noted agents may be administered as immediate
30 release, or as extended release dosage forms, either together with a suitable CSAID compound, or separately. The compositions may be administered sequentially, in combination with, or contemporaneously with a CSAID agent. The administration route of the second agent may also differ from that of the CSAID agent, and hence the dosing schedule may vary accordingly.

35 Cetirizine HCl manufacture and dosing is described in US Patent 4,525,358; fexofenadine manufacture and dosing is described in US Patents 4,524,129; US 5,375,693; US 5,578,610; US 5,855,912; US 5,932,247; and US 6,037,353.

WO 01/19322

PCT/US00/25386

Loratadine and DCL manufacture and dosing are described in US patent 4,282,233; US 4,371,516; US 4,659,716; US 4,863,931; US 5,314,697; and US 5,595,997.

Zamanivar dosing is disclosed in US Patents 4,627,432; US 4,778,054; US 4,811,731; US 5,035,237; US 5,360,817; and US 5,648,379. Oseltamivir dosing is disclosed in US Patents US 5,763,483; US 5,866,601; and US 5,952,375.

The CSPB/p38 inhibitor may be administered systemically or non-systemically, such as orally, buccally, topically (intranasal) or via inhalation (aerosol), or both topically and via inhalation. As noted above, the second therapeutic agent may be administered by any suitable means, including parenteral, suppository, etc. which means of administration is not necessarily by the same route, nor concurrent therewith.

As used herein "topically" shall include non-systemic administration. This includes the application of a compound externally to the epidermis or the buccal cavity and/or the instillation of such a compound into the ear, eye and nose.

As used herein "systemic administration" refers to oral, intravenous, intraperitoneal and intramuscular administration, subcutaneous intranasal, intrarectal, or intravaginal.

It will be recognized by one of skill in the art that the optimal quantity and spacing of individual dosages of a CSBP/p38 inhibitor will be determined by the nature and extent of the condition being treated, the form, route and site of administration, and the particular patient being treated, and that such optimums can be determined by conventional techniques. It will also be appreciated by one of skill in the art that the optimal course of treatment, i.e., the number of doses of a CSBP/p38 inhibitor given per day for a defined number of days, can be ascertained by those skilled in the art using conventional course of treatment determination tests.

Methods:

Cell lines, rhinovirus serotype 39, and influenza virus A/PR/8/34 were purchased from American Type Culture Collection (ATCC). BEAS-2B cells were cultured according to instructions provided by ATCC using BEGM (bronchial epithelial growth media) purchased from Clonetics Corp. HELA cell cultures, used for detection and titration of virus, were maintained in Eagle's minimum essential media containing 10% fetal calf serum, 2mM l-glutamine, and 10 mM HEPES buffer (MEM).

A modification of the method reported by Subauste et al., *Supra*, for in vitro infection of human bronchial epithelial cells with rhinovirus was used in these studies. BEAS-2B cells (2×10^5 /well) were cultured in collagen-coated wells for 24 hours prior to infection with rhinovirus. Rhinovirus serotype 39 was added to cell

WO 01/19322

PCT/US00/25386

cultures for one hour incubation at 34°C after which inoculum was replaced with fresh media and cultures were incubated for an additional 72 hours at 34°C. Supernatants collected at 72 hours post-infection were assayed for cytokine protein concentration by ELISA using commercially available kits (R&D Systems). Virus yield was also determined from culture supernatants using a microtitration assay in HELA cell cultures (*Subauste et al., supra 1995*). In cultures treated with p38 kinase inhibitors, drug was added 30 minutes prior to infection. Stocks of compounds were prepared in DMSO (10 mM drug) and stored at -20°C.

For detection of p38 kinase, cultures were incubated in basal media without growth factors and additives to reduce endogenous levels of activated p38 kinase. Cells were harvested at various timepoints after addition of rhinovirus. Detection of tyrosine phosphorylated p38 kinase by immunoblot was analyzed by a commercially available kit and was performed according to the manufacturer's instructions (PhosphoPlus p38 MAPK Antibody Kit: New England BioLabs Inc.).

In some experiments, BEAS-2B cells were infected with influenza virus (strain A/PR/8/34) in place of rhinovirus. Culture supernatant was harvested 48 and 72 hour post-infection and tested by ELISA for cytokine as described above.

Cells and Virus: Influenza A/PR/8/34 sub type H1N1 (VR-95 American Type Culture Collection, Rockville, MD) was grown in the allantoic cavity of 10 day old chicken eggs. Following incubation at 37°C, and refrigeration for 2 1/2 hours at 4°C, allantoic fluid was harvested, pooled, and centrifuged (1,000 rcf; 15 min; 4°C) to remove cells. Supernatant was aliquoted and stored at -70°C. The titer of the stock culture of virus was 1.0×10^{10} Tissue Culture Infective Dose/ml (TCID₅₀)

Inoculation procedure: Four-six week old female Balb/cAnNcr1Br mice were obtained from Charles River, Raleigh, NC. Animals were infected intranasally. Mice were anesthetized by intraperitoneal injection of Ketamine (40mg/kg; Fort Dodge Labs, Fort Dodge, Ia) and Xylazine (5 mg/kg; Miles, Shawnee Mission, Ks) and then inoculated with 100 TCID₅₀ of PR8 diluted in PBS in 20 ul. Animals were observed daily for signs of infection. All animal studies were approved by SmithKline Beecham Pharmaceuticals Institutional Animal Care and Use Committee.

Virus titration: At various times post infection, animals were sacrificed and lungs were aseptically harvested. Tissues were homogenized, in vials containing 1 micron glass beads (Biospec Products, Bartlesville, OK) and 1 ml. of Eagles minimal essential medium. Cell debris was cleared by centrifugation at 1,000 rcf for 15 minutes at 4°C, and supernatants were serially diluted on Madin-Darby canine kidney (MDCK) cells. After 5 days of incubation at 37°C (5% CO₂), 50 µl of 0.5%

chick red blood cells were added per well, and agglutination was read after 1 hour at room temperature. The virus titer is expressed as 50% tissue culture infective dose (TCID₅₀) calculated by logistic regression.

ELISA: Cytokine levels were measured by quantitative ELISA using commercially available kits. Ear samples were homogenized using a tissue minser in PBS. Cell debris was cleared by centrifugation at 14,000 rpm for 5 minutes. The cytokine concentrations and thresholds were determined as described by the manufacturer; IL-6, IFN- γ , and KC (R&D Systems, Minneapolis, MN).

Myeloperoxidase Assay: Myeloperoxidase (MPO) activity was determined kinetically as described by Bradley et al. (1982). Briefly, rabbit cornea were homogenized in Hexadecyl Trimethyl-Ammonium Bromide (HTAB) (Sigma Chemical Co. St. Louis, Mo) which was dissolved in 0.5 M Potassium phosphate buffer (J.T. Baker Scientific, Phillipsburg, NJ). Following homogenization, the samples were subjected to freeze-thaw-sonication (Cole-Parmer 8853, Cole-Parmer, Vernon Hills, IL) 3 times.

Suspensions were then cleared by centrifugation at 12,500 x g for 15 minutes at 4°C. MPO enzymatic activity was determined by colorimetric change in absorbance during a reaction of O-Dianisidine dihydrochloride (ODI) 0.175 mg/ml (Sigma Chemical Co. St. Louis, Mo) with .0002% Hydrogen peroxide (Sigma Chemical Co. St. Louis, Mo). Measurements were performed by using a Beckman Du 640 Spectrophotometer (Fullerton, Ca.) fitted with a temperature control device. 50 μ l of material to be assayed was added to 950 μ l of ODI and change in absorbance was measured at a wave length of 460 nm for 2 minutes at 25°C.

Whole Body Plethysmography: Influenza virus infected mice were placed into a whole body plethysmograph box with an internal volume of approximately 350-ml. A bias airflow of one l/min was applied to the box and flow changes were measured and recorded with a Buxco XA data acquisition and respiratory analysis system (Buxco Electronics, Sharon, CT). Animals were allowed to acclimate to the plethysmograph box for 2 min. before airflow data was recorded. Airway measurements were calculated as Penh (enhanced pause). Penh has previously been shown as an index of airway obstruction and correlates with increased intrapleural pressure. The algorithm for Penh calculation is as follows: $\text{Penh} = [(\text{expiratory time} / \text{relaxation time}) - 1] \times (\text{peak expiratory flow} / \text{peak inspiratory flow})$ where relaxation time is the amount of time required for 70% of the tidal volume to be expired.

Determination of arterial oxygen saturation. A Nonin veterinary hand held pulse oximeter 8500V with lingual sensor (Nonin Medical, Inc., Plymouth MN) was used

to determine daily arterial oxygen saturation %SpO₂ as described (Sidwell et al. 1992 Antimicrobial Agents and Chemotherapy 36:473-476).

Results:

5 *Inhibition of cytokine production by specific inhibitors of p38 MAP kinase:*

Consistent with published reports, IL-6, IL-8, and GM-CSF were detected 72 hours post-infection of BEAS-2B cells with rhinovirus-39 (multiplicity of infection; MOI 1.0) (figure 1). Production of IL-6, IL-8, and GM-CSF was not mediated through IL-1 or TNF produced in response to rhinovirus infection since addition of
10 neutralizing antibodies to IL-1 and TNF to the infected cultures did not reduce the amount of IL-6, IL-8 or GM-CSF produced (not shown). Productive infection of cells was confirmed by titering infectious supernatants from BEAS-2B cells on HELA monolayers. There was low but consistent replication of virus during the 72 hour culture period resulting in $1.22 \pm 0.3 \log_{10}$ TCID₅₀ increase over the initial
15 input inoculum (n=6 experiments). To investigate the role of p38 kinase signal transduction in rhinovirus-induced cytokine production by epithelial cells, specific p38 kinase inhibitors SB203580, Compound II, Compound III, and an inactive analog, SKF106978, were tested for their ability to inhibit cytokine production in rhinovirus-infected BEAS-2B cell cultures. The compound (4-Fluorophenyl)-2-(4-methylsulfinylphenyl)-5-(4-pyridyl)-imidazole is alternatively referred to as SB
20 203580 and may be found in US patent 5,656,644. The compound *trans*-1-(4-Hydroxycyclohexyl)-4-(4-fluorophenyl)-5-[(2-methoxy)pyrimidin-4-yl]imidazole also known as Compound II may be found in WO 97/25048. The compound 4-(4-Fluorophenyl)-5-[(2-phenylamino)pyrimidin-4-yl]-1-(piperidin-4-yl)imidazole, also
25 known as Compound III may be found in US Patent 5,658,903. The compound 2-(4-Methylsulfinyl)-3-[4-(2-methylpyridyl)-6,7-dihydro[5H]pyrrolo[1,2-a]imidazole, is also known as SB 106978. Concentrations of IL-8, IL-6 and GM-CSF in culture supernatants from infected cells treated with inhibitors were all lower than those from untreated infected cultures (figure 2). IL-6 was the most sensitive to inhibition
30 with significant inhibition (40%) being observed with SB 203580 concentrations as low as 30 nM. GM-CSF was the least sensitive to inhibition by SB 203580, with an IC₅₀ of approximately 4 uM.

Another inhibitor of p38 kinase, Compound II, was slightly more potent in inhibiting GM-CSF with an IC₅₀ of approximately 1 uM. Compound II was
35 comparable to SB 203580 in inhibiting IL-6 and IL-8 production. As expected based on the relative potency of these compounds in specific binding to p38 kinase (data shown in legend box), cytokine inhibition was greatest with Compound III,

WO 01/19322

PCT/US00/25386

with an IC₅₀ value < 10 nM for IL-6, while SKF106978 was inactive at all concentrations tested. Maximum effect obtained by any of the p38 kinase inhibitors against any of the three cytokines was 50% -70% inhibition. The inhibition of cytokine production by CSAIDs was not due to general cell cytotoxicity as determined by standard XTT assays (CC₅₀ > 40 uM for all compounds tested) (Roehm et al., J. of Immunol. Methods 142:257-265 (1991)).

These compounds also did not exhibit direct antiviral activity as assessed using a standard HELA cell antiviral assay (MIC₅₀ > 10 uM for all compounds tested) (Andries et al., Journal of Virology 64(3):1117-1123 (1990) or by measuring virus yield in the RV-infected BEAS-2B cultures directly (not shown).

Activation of p38 kinase by rhinovirus infection:

The presence of tyrosine phosphorylated p38 kinase was measured by immunoblot at various times after the addition of virus to BEAS-2B cultures. Rhinovirus infection of BEAS-2B cells resulted in an increase in phosphorylated p38 kinase that was both dose and time-dependent. Increases in phosphorylated p38 kinase were evident by 15 minutes post exposure to rhinovirus-39 (MOI 10), appeared to peak by 30 minutes after addition of virus and remained elevated 60 minutes post-infection (figure 3). In addition, rhinovirus-induced tyrosine phosphorylation of p38 kinase was dose-dependent (figure 4). When cells were cultured in the absence of virus, there was no increase in the amount of tyrosine phosphorylation of p38 kinase at any of the timepoints tested. Overall levels of p38 kinase protein were comparable between all the groups indicating that virus infection caused phosphorylation of p38 kinase without de novo synthesis of protein (figures 3 and 4).

Effects on in vitro influenza virus infection:

Exposure of BEAS-2B cells with influenza virus (A/PR/8/34; MOI 1.0) also resulted in elaboration of IL-8 and IL-6 as measured 48 - 72 hours post-infection, although the secreted protein levels were lower than that obtained with rhinovirus infection. Consistent with observations in rhinovirus-infected cells, treatment of influenza infected-BEAS-2B cells with p38 kinase inhibitors, Compound IV, 1-(4-Piperidinyl)-4-(4-fluorophenyl)-5-[(2-methylphenyl)amino]pyrimidin-4-yl]imidazole and Compound II, 1-*trans*-4-hydroxycyclohexyl)-4-(4-fluorophenyl)-5-[(2-methoxy)pyrimidin-4-yl]imidazole, was also effective in inhibiting IL-6 and IL-8 production.

Effects on in vivo influenza virus infection:

Five (5) independent and reproducible studies demonstrated the efficacy of therapeutic dosing with Compound V, 1-(4-Piperidinyl)-4-(4-fluorophenyl)-5-(2-methoxy-4-pyrimidinyl)imidazole and Compound VI, 1-(1,3-Dihydroxyprop-2-yl)-4-(4-fluorophenyl)-5-[2-phenoxy-4-pyrimidin-4-yl]imidazole at improving clinical disease in the murine influenza pneumonia model. BALB/c mice were dosed orally b.i.d. on days 3-8 post influenza A/PR8 and monitored daily for weight loss, pulmonary functions and arterial blood oxygen levels %SpO₂. The antiviral Tamiflu was used as control and demonstrated 47% improvement in pulmonary functions (p<0.01 days 5-12), 64% improved %SpO₂ (p<0.01 days 5-18), and prevention of weight loss relative to placebo. The optimal dose of Compound VI was 10 mg/kg leading to 39% improvement in pulmonary functions (p<0.01 days 5-12), 30% improvement in %SpO₂ (p<0.01 days 5-13, p<0.05 days 14-15), and a similar effect on weight loss as Tamiflu treatment. Efficacy was observed in doses as low as 1 mg/kg: 27% improvement in pulmonary functions (p<0.01 days 6-9), 11.6% (p<0.01 Days 7-13). At 0.1 mg/kg, we observed 19% improvement in pulmonary functions (p<0.05 day 7,8) but no effect on %SpO₂ or weight loss. At 10 mg/kg, Compound VI was equally effective to Compound V at 30 mg/kg. Samples were collected for evaluation of virus titers and cytokines in lung homogenates. A non-significant trend for inhibition of lung cytokines IL-6, KC, IFN-gamma, and RANTES was observed. There was no negative effect on lung virus titers.

No negative effect on immunity to secondary influenza virus infection: In two studies, mice treated with Compound VI or Compound V during acute PR/8 (H1N1) influenza infection were protected from a lethal challenge with the same virus as demonstrated by 100% survival and normal pulmonary functions. All control primary infection animals died by Day 7. Thus, the CSAIDS have no effect on immunity to a homologous challenge.

All publications, including but not limited to patents and patent applications, cited in this specification are herein incorporated by reference as if each individual publication were specifically and individually indicated to be incorporated by reference herein as though fully set forth.

The above description fully discloses the invention including preferred embodiments thereof. Modifications and improvements of the embodiments specifically disclosed herein are within the scope of the following claims. Without

WO 01/19322

PCT/US00/25386

further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. Therefore the Examples herein are to be construed as merely illustrative and not a limitation of the scope of the present invention in any way. The embodiments of the invention in
5 which an exclusive property or privilege is claimed are defined as follows.